

Human TGF-β1 (CHO) premium grade

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1. Description

Products

Human TGF- β 1 (CHO), premium grade. Recombinant human transforming growth factor β 1.

Content in µg	Order no.
5	130-126-723
25	130-126-721
100	130-126-724
1000, liquid	130-126-722

Biological activity

The ED₅₀ is \leq 0.2 ng/mL corresponding to an activity of \geq 5×10⁶ U/mg. Lot-specific activities are stated in the Certificate of Analysis (www. miltenyibiotec.com/certificates).

▲ Note: The ED50 is determined by inhibition assay using IL-5 induced TF-1 cells according to Randall $\it et al.$ The proliferation assay was calibrated with the standard for human TGF- $\it β1$ (NIBSC code 89/514) provided by the WHO/National Institute for Biological Standards and Control.

Primary structure

Two identical, non-glycosylated disulfidelinked polypeptide chains (112 amino acid residues without propeptide LAP).

Molecular mass

25.6 kDa (dimer).

Source

Produced in CHO cells.

Product format

 $5 \mu g$, $25 \mu g$, $100 \mu g$: Lyophilized from a filtered (0.2 μm) buffer solution.

1000 um Liquid filtored (0.2 um

 $1000~\mu g$: Liquid, filtered (0.2 μm) phosphate buffer solution without stabilizers.

Stabilizer Purity Mannitol and trehalose (5 $\mu g, 25~\mu g, 100~\mu g).$ >97% as determined by SDS-PAGE analysis.

Endotoxin level

Low endotoxin (<0.1 EU/ μ g cytokine) as determined by Limulus Amebocyte Lysate (LAL) assay.

Storage

Lyophilized Human TGF- β 1 (CHO), premium grade should be stored at $-20\,^{\circ}$ C. The expiration date is indicated on the vial label. Upon reconstitution aliquots should be stored at $-20\,^{\circ}$ C or below. Avoid repeated freeze-thaw cycles.

Reconstitution

It is recommended to reconstitute lyophilized Human TGF- β 1 (CHO), premium grade with deionized sterile-filtered water to a final concentration of 0.1–1.0 mg/mL in a minimal volume of 250 μ L. Further dilutions should be prepared with 0.1% bovine serum albumin (BSA) or human serum albumin (HSA) in phosphate-buffered saline.

1.1 Background information

Transforming growth factor β1 (TGF-β1) belongs to a family of homologous, disulfide-linked, homodimeric proteins. These highly pleiotropic cytokines inhibit proliferation of most cells but can promote the growth of mesenchymal cells and enhance extracellular matrix formation. The pivotal function of TGF-β1 in the immune system is to mediate immunosuppression and maintain tolerance by regulating lymphocyte proliferation, differentiation, and survival. In addition, TGF-β1 controls inflammatory responses through chemotactic attraction and activation of inflammatory cells and fibroblasts. TGF-\(\beta\)1 is produced by many cell types but is reported to be most abundant in mammalian platelets and bone. All three TGF- β members are synthesized as a homodimeric precursor of 390 residues, which is intracellularly processed by proteolysis into a 112 aa form. The resulting N-terminal latency associated peptide (LAP) remains non-covalently associated with the TGF- β dimer, and the complex binds to another protein called latent TGF-β-binding protein (LTBP), forming a larger complex called Large Latent Complex (LLC). The LLC is secreted into the extracellular matrix, and prevents the binding of TGF-β to its specific cell surface receptor. Several extracellular factors such as matrix metalloproteases, low pH, reactive oxygen species and thrombospondin-1 can induce release of the active mature TGF- β dimer from the inactive complex. This sophisticated mechanism of activation is important for a fine-tuning of TGF-β signaling. Human TGF-β1 is a recombinant homodimer corresponding to the fully mature form of TGF-\$1 without LAP. The amino acid sequence of human TGF-β1 shares 99% identity with TGF-β1 from mouse and rat, therefore human TGF- $\beta 1$ is commonly used also for mouse cell culture.

1.2 Applications

Human TGF- $\beta 1$ (CHO) can be used for a variety of applications, including:

- In vitro differentiation of naive CD4⁺ T cells towards Th17 cells.
- In vitro generation of FoxP3⁺ inducible regulatory T cells (iTregs).
- Embryonic stem cell differentiation models, for example, for vasculogenesis and angiogenesis.
- In vitro chondrogenesis of mesenchymal progenitor cells and redifferentiation of expanded chondrocytes.

Optimal concentration for a specific application should be determined by a dose-response experiment.

2. References

1. Randall, L. A. $\it{et~al.}$ (1993) A novel, sensitive bioassay for transforming growth factor $\it{\beta}$. J. Immunol. Methods 164: 61–67.

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