

## Report

# T<sub>H</sub>17 cell differentiation

## Effective and reliable *in vitro* generation of human T<sub>H</sub>17 cells

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## Introduction

T<sub>H</sub>17 cells are a unique lineage of CD4<sup>+</sup> T helper cells, which characteristically secrete IL-17 at high levels and, to a lesser extent, IL-22. T<sub>H</sub>17 cells are involved in immune responses against extracellular pathogens, such as *Candida*<sup>1</sup>.

T<sub>H</sub>17 cells also play a critical role in the pathogenesis of autoimmune diseases and other atopic and chronic inflammatory disorders<sup>2</sup>.

T<sub>H</sub>17 cell differentiation is orchestrated by multiple cytokines. Naive human T helper cells can be induced to differentiate *in vitro* into T<sub>H</sub>17 cells by exposure to TGF- $\beta$  and proinflammatory cytokines, particularly IL-1 $\beta$  and IL-6, whereas IL-23 promotes survival and proliferation of T<sub>H</sub>17 cells<sup>3,4</sup>.

The possibility of generating a highly enriched, consistent population of T<sub>H</sub>17 cells *in vitro* provides opportunities to investigate the biology of T<sub>H</sub>17 cells in detail.

## Materials and methods

### Recombinant cytokines and neutralizing antibodies

T<sub>H</sub>17-polarizing cytokines, i.e., human IL-1 $\beta$ , IL-6, IL-23, and TGF- $\beta$ 1, from Miltenyi Biotec (MACS<sup>®</sup> Cytokines) were tested against the same cytokines from other providers. Functional-grade antibodies against IFN- $\gamma$  (clone 45-15) and IL-4 (clone 7A3-3) were from Miltenyi Biotec.

### Isolation of naive human T cells

Human PBMCs were prepared from peripheral blood of healthy donors by density gradient centrifugation.

Naive CD4<sup>+</sup>CD45RA<sup>+</sup> T cells were purified from PBMCs using the Naive T Cell Isolation Kit, human (Miltenyi Biotec).

### T<sub>H</sub>17 cell generation

Naive CD4<sup>+</sup> T cells were seeded in 24-well plates (1 $\times$ 10<sup>6</sup> per well) and cultured in a serum-free medium. Cells were stimulated by using the T Cell Activation/Expansion Kit, human (Miltenyi Biotec), which is based on MACSiBead<sup>™</sup> Particles loaded with CD2, CD3, and CD28 antibodies. The bead-to-cell ratio amounted to 1:2. Cells were cultured in the presence of the T<sub>H</sub>17-polarizing cytokines, IL-1 $\beta$  (20 ng/mL), IL-6 (30 ng/mL), IL-23 (30 ng/mL), and TGF- $\beta$ 1 (2.25 ng/mL), in addition to Anti-IFN- $\gamma$  (1  $\mu$ g/mL) and Anti-IL-4 (2.5  $\mu$ g/mL) antibodies. Cells were cultured for 7 days at 37 °C, in an atmosphere of 5% CO<sub>2</sub>, without any media exchange. Enzyme-linked immunosorbent assays (ELISA) were performed as described.

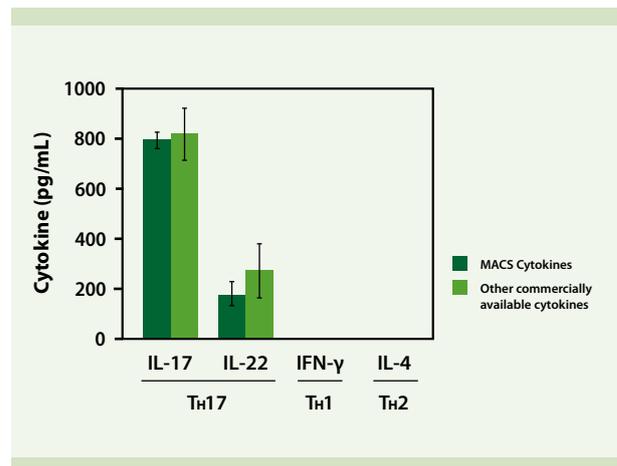
### ELISA

Cell culture supernatants were collected, filtered, and measured for their content of IFN- $\gamma$ , IL-4, IL-17, IL-22, by using ELISA (R&D Systems) according to the manufacturer's protocol.

## Results and conclusion

Here we show that a combination of recombinant human TGF- $\beta$ 1, IL-23, and the proinflammatory cytokines IL-1 $\beta$  and IL-6 effectively drives differentiation of isolated naive human CD4<sup>+</sup> T cells into T<sub>H</sub>17 cells. In addition to IL-17 production, the hallmark of T<sub>H</sub>17 cells, these T<sub>H</sub>17-polarizing conditions also induced low-level production of IL-22 (fig. 1). By contrast, IFN- $\gamma$  and IL-4, which are characteristic of T<sub>H</sub>1 and T<sub>H</sub>2 cells, respectively, could not be detected.

The quality of cell culture ingredients dramatically influences cell behavior. Consistent, high-quality products are essential for reliable and functionally relevant cell culture results. We compared recombinant cytokines from different providers. Premium-grade cytokines from Miltenyi Biotec showed at least equal performance compared to other commercially available cytokines and consistently generated T<sub>H</sub>17 cells *in vitro* under the reported T<sub>H</sub>17-polarizing conditions.



**Figure 1:** Naive human CD4<sup>+</sup> T cells were stimulated with MACS<sup>i</sup>Bead Particles loaded with CD2, CD3, and CD28 antibodies, in the presence of a T<sub>H</sub>17-polarizing cytokine cocktail (IL-1 $\beta$ , IL-6, IL-23, and TGF- $\beta$ 1), in serum-free medium. Concentrations of IL-17, IL-22, IFN- $\gamma$ , and IL-4 in cell culture supernatants were determined by ELISA on day 7 (n=2, independent donors). Error bars represent SEM.

### References

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