

Contents

1. Description
 - 1.1 Principle of the MACS® Separation
 - 1.2 Background information
 - 1.3 Applications
 - 1.4 Reagent and instrument requirements
2. Protocol
 - 2.1 Sample preparation
 - 2.2 Magnetic labeling
 - 2.3 Magnetic separation
3. Example of a separation using the Anti-HLA-DR MicroBeads

1. Description

Components	2 mL Anti-HLA-DR MicroBeads, human: MicroBeads conjugated to monoclonal mouse anti HLA-DR antibodies (isotype: mouse IgG1).
Capacity	For 10 ⁹ total cells, up to 100 separations.
Product format	Anti-HLA-DR MicroBeads are supplied in buffer containing stabilizer and 0.05% sodium azide.
Storage	Store protected from light at 2–8 °C. Do not freeze. The expiration date is indicated on the vial label.

1.1 Principle of the MACS® Separation

First, the HLA-DR⁺ cells are magnetically labeled with Anti-HLA-DR MicroBeads. Then the cell suspension is loaded onto a MACS® Column which is placed in the magnetic field of a MACS Separator. The magnetically labeled HLA-DR⁺ cells are retained on the column. The unlabeled cells run through; this cell fraction is thus depleted of HLA-DR⁺ cells. After removing the column from the magnetic field, the magnetically retained HLA-DR⁺ cells can be eluted as the positively selected cell fraction.

1.2 Background information

Anti-HLA-DR MicroBeads have been developed for the separation of cells based on the expression of the HLA-DR antigen. HLA-DR is expressed on dendritic cells, B cells, monocytes, macrophages, activated T cells, activated NK cells, hematopoietic progenitor cells, and some epithelial cells.

1.3 Applications

- Positive selection or depletion of antigen-presenting cells, e.g., B cells, monocytes, and dendritic cells, from human PBMCs (peripheral blood mononuclear cells) or lymphoid tissue.
- Isolation of primitive CD34⁺ HLA-DR⁻ hematopoietic progenitor cells in combination with CD34 MultiSort Kit (# 130-056-701) from human peripheral blood, cord blood, or bone marrow.
- Positive selection of HLA-DR expressing cells from human PBMCs or buffy coat for serological typing of HLA-DR class II molecules.

1.4 Reagent and instrument requirements

- **Buffer:** Prepare a solution containing phosphate-buffered saline (PBS), pH 7.2, 0.5% bovine serum albumin (BSA), and 2 mM EDTA by diluting MACS BSA Stock Solution (# 130-091-376) 1:20 with autoMACS® Rinsing Solution (# 130-091-222). Keep buffer cold (2–8 °C). Degas buffer before use, as air bubbles could block the column.

▲ **Note:** EDTA can be replaced by other supplements such as anticoagulant citrate dextrose formula-A (ACD-A) or citrate phosphate dextrose (CPD). BSA can be replaced by other proteins such as human serum albumin, human serum, or fetal bovine serum (FBS). Buffers or media containing Ca²⁺ or Mg²⁺ are not recommended for use.

- **MACS Columns and MACS Separators:** HLA-DR⁺ cells can be enriched by using MS, LS, or XS Columns or depleted with the use of LD Columns. Cells that strongly express the HLA-DR antigen can also be depleted using MS, LS, or XS Columns. Positive selection or depletion can also be performed by using the autoMACS Pro or the autoMACS Separator.

Column	Max. number of labeled cells	Max. number of total cells	Separator
Positive selection			
MS	10 ⁷	2×10 ⁸	MiniMACS, OctoMACS, VarioMACS, SuperMACS II
LS	10 ⁸	2×10 ⁹	MidiMACS, QuadroMACS, VarioMACS, SuperMACS II
XS	10 ⁹	2×10 ¹⁰	SuperMACS II
Depletion			
LD	10 ⁸	5×10 ⁸	MidiMACS, QuadroMACS, VarioMACS, SuperMACS II
Positive selection or depletion			
autoMACS	2×10 ⁸	4×10 ⁹	autoMACS Pro, autoMACS

▲ **Note:** Column adapters are required to insert certain columns into the VarioMACS™ or SuperMACS™ II Separators. For details refer to the respective MACS Separator data sheet.

- (Optional) Fluorochrome-conjugated Anti-HLA-DR antibodies for flow cytometric analysis, e.g., Anti-HLA-DR-PE (# 130-095-298) or Anti-HLA-DR-APC (# 130-095-297). For more information about antibodies refer to www.miltenyibiotec.com/antibodies.

- (Optional) Propidium Iodide Solution (# 130-093-233) or 7-AAD for flow cytometric exclusion of dead cells.
- (Optional) Dead Cell Removal Kit (# 130-090-101) for the depletion of dead cells.
- (Optional) Pre-Separation Filters, 30 μm (# 130-041-407) to remove cell clumps.

2. Protocol

2.1 Sample preparation

When working with anticoagulated peripheral blood or buffy coat, peripheral blood mononuclear cells (PBMCs) should be isolated by density gradient centrifugation, for example, using Ficoll-Paque™.

▲ **Note:** To remove platelets after density gradient separation, resuspend cell pellet in buffer and centrifuge at 200 \times g for 10–15 minutes at 20 °C. Carefully aspirate supernatant. Repeat washing step.

When working with tissues or lysed blood, prepare a single-cell suspension using standard methods.

For details refer to the protocols section at www.miltenyibiotec.com/protocols.

▲ Dead cells may bind non-specifically to MACS MicroBeads. To remove dead cells, we recommend using density gradient centrifugation or the Dead Cell Removal Kit (# 130-090-101).



2.2 Magnetic labeling

▲ Work fast, keep cells cold, and use pre-cooled solutions. This will prevent capping of antibodies on the cell surface and non-specific cell labeling.

▲ Volumes for magnetic labeling given below are for up to 10^7 total cells. When working with fewer than 10^7 cells, use the same volumes as indicated. When working with higher cell numbers, scale up all reagent volumes and total volumes accordingly (e.g. for 2×10^7 total cells, use twice the volume of all indicated reagent volumes and total volumes).

▲ For optimal performance it is important to obtain a single-cell suspension before magnetic labeling. Pass cells through 30 μm nylon mesh (Pre-Separation Filters, 30 μm , # 130-041-407) to remove cell clumps which may clog the column. Moisten filter with buffer before use.

▲ The recommended incubation temperature is 2–8 °C. Higher temperatures and/or longer incubation times may lead to non-specific cell labeling. Working on ice may require increased incubation times.

Magnetic labeling of human PBMCs or cells from lymphoid tissue

1. Determine cell number.
2. Centrifuge cell suspension at 300 \times g for 10 minutes. Aspirate supernatant completely.
3. Resuspend cell pellet in 80 μL of buffer per 10^7 total cells.
4. Add 20 μL of Anti-HLA-DR MicroBeads per 10^7 total cells.
5. Mix well and incubate for 15 minutes in the refrigerator (2–8 °C).

6. (Optional) Add staining antibodies, e.g., 10 μL of Anti-HLA-DR-PE (# 130-095-298), and incubate for 5 minutes in the dark in the refrigerator (2–8 °C).
7. Wash cells by adding 1–2 mL of buffer per 10^7 cells and centrifuge at 300 \times g for 10 minutes. Aspirate supernatant completely.
8. Resuspend up to 10^8 cells in 500 μL of buffer.
 - ▲ **Note:** For higher cell numbers, scale up buffer volume accordingly.
 - ▲ **Note:** For depletion with LD Columns, resuspend up to 1.25×10^8 cells in 500 μL of buffer.
9. Proceed to magnetic separation (2.3).

Magnetic labeling of cells from buffy coat

1. Centrifuge 2–3 mL anticoagulated peripheral blood at 400 \times g for 35 minutes. Carefully collect leukocyte-enriched interphase (buffy coat) in 300 μL volume.
 - ▲ **Note:** For lysis of erythrocytes refer to 4. Appendix.
2. Add 60 μL of Anti-HLA-DR MicroBeads per 300 μL of buffy coat.
3. Mix well and incubate for 20 minutes in the refrigerator (2–8 °C).
4. (Optional) Add staining antibodies, e.g., 10 μL of Anti-HLA-DR-PE (# 130-095-298), and incubate for 5 minutes in the dark in the refrigerator (2–8 °C).
5. Wash cells by adding 1–2 mL of buffer per 10^7 cells and centrifuge at 300 \times g for 10 minutes. Aspirate supernatant completely.
6. Resuspend up to 10^8 cells in 500 μL of buffer.
 - ▲ **Note:** For higher cell numbers, scale up buffer volume accordingly.
 - ▲ **Note:** For depletion with LD Columns, resuspend up to 1.25×10^8 cells in 500 μL of buffer.

Proceed to magnetic separation (2.3).



2.3 Magnetic separation

▲ Choose an appropriate MACS Column and MACS Separator according to the number of total cells and the number of HLA-DR⁺ cells. For details refer to the table in section 1.4.

▲ Always wait until the column reservoir is empty before proceeding to the next step.

Magnetic separation with MS or LS Columns

1. Place column in the magnetic field of a suitable MACS Separator. For details refer to the respective MACS Column data sheet.
2. Prepare column by rinsing with the appropriate amount of buffer:

MS: 500 μL	LS: 3 mL
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3. Apply cell suspension onto the column. Collect flow-through containing unlabeled cells.

- Wash column with the appropriate amount of buffer. Collect unlabeled cells that pass through and combine with the flow-through from step 3.

MS: 3×500 μ L LS: 3×3 mL

▲ **Note:** Perform washing steps by adding buffer aliquots only when the column reservoir is empty.

- Remove column from the separator and place it on a suitable collection tube.
 - Pipette the appropriate amount of buffer onto the column. Immediately flush out the magnetically labeled cells by firmly pushing the plunger into the column.
- MS: 1 mL LS: 5 mL
- (Optional) To increase the purity of HLA-DR⁺ cells, the eluted fraction can be enriched over a second MS or LS Column. Repeat the magnetic separation procedure as described in steps 1 to 6 by using a new column.

Magnetic separation with XS Columns

For instructions on the column assembly and the separation refer to the XS Column data sheet.

Depletion with LD Columns

- Place LD Column in the magnetic field of a suitable MACS Separator. For details refer to the LD Column data sheet.
- Prepare column by rinsing with 2 mL of buffer.
- Apply cell suspension onto the column.
- Collect unlabeled cells that pass through and wash column with 2×1 mL of buffer. Collect total flow-through; this is the unlabeled cell fraction. Perform washing steps by adding buffer two times. Only add new buffer when the column reservoir is empty.

Magnetic separation with the autoMACS[®] Pro Separator or the autoMACS[®] Separator

▲ Refer to the respective user manual for instructions on how to use the autoMACS[®] Pro Separator or the autoMACS Separator.

▲ Buffers used for operating the autoMACS Pro Separator or the autoMACS Separator should have a temperature of ≥ 10 °C.

▲ Program choice depends on the isolation strategy, the strength of magnetic labeling, and the frequency of magnetically labeled cells. For details refer to the section describing the cell separation programs in the respective user manual.

Magnetic separation with the autoMACS[®] Pro Separator

- Prepare and prime the instrument.
- Apply tube containing the sample and provide tubes for collecting the labeled and unlabeled cell fractions. Place sample tube in row A of the tube rack and the fraction collection tubes in rows B and C.
- For a standard separation choose one of the following programs:

Positive selection: Possel

Collect positive fraction in row C of the tube rack.

Depletion: Depletes

Collect negative fraction in row B of the tube rack.

Magnetic separation with the autoMACS[®] Separator

- Prepare and prime the instrument.
- Apply tube containing the sample and provide tubes for collecting the labeled and unlabeled cell fractions. Place sample tube at the uptake port and the fraction collection tubes at port neg1 and pos1.
- For a standard separation choose one of the following programs:

Positive selection: Possel

Collect positive fraction from outlet pos1.

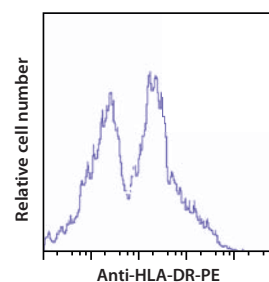
Depletion: Depletes

Collect negative fraction from outlet port neg1.

3. Example of a separation using the Anti-HLA-DR MicroBeads

HLA-DR⁺ cells were isolated from human PBMCs using Anti-HLA-DR MicroBeads, an MS Column, and a MiniMACS[™] Separator and analyzed by flow cytometry using the MACSQuant[®] Analyzer. Cell debris and dead cells were excluded from the analysis based on scatter signals and propidium iodide fluorescence.

Original fraction is shown as purple line, positive fraction containing cells selected with Anti-HLA-DR MicroBeads are shown as a black line.



All protocols and data sheets are available at www.miltenyibiotec.com.

Warnings

Reagents contain sodium azide. Under acidic conditions sodium azide yields hydrazoic acid, which is extremely toxic. Azide compounds should be diluted with running water before discarding. These precautions are recommended to avoid deposits in plumbing where explosive conditions may develop.

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